

Carbonic anhydrase inhibition. Insight into the characteristics of zonisamide, topiramate, and the sulfamide cognate of topiramate

RICHARD P. SHANK[†], VIRGINIA L. SMITH-SWINTOSKY[‡], & BRUCE E. MARYANOFF^{*}

Research & Early Development, Johnson & Johnson Pharmaceutical Research & Development, Spring House, Pennsylvania 19477-0776, USA

(Received 2 March 2007; accepted 25 May 2007)

Abstract

Some useful therapeutic agents inhibit certain carbonic anhydrase (CA) isozymes to varying degrees. We have conducted enzyme kinetics studies in a 4-nitrophenyl acetate (4-NPA) hydrolysis assay with the marketed antiepileptic drugs topiramate (1) and zonisamide (2) to determine if their full inhibition of human CA-II and CA-I requires extended preincubation conditions. We found that neither 1 nor 2 requires appreciable preincubation with either enzyme to manifest full inhibitory activity. We also examined the sulfamide cognate of topiramate (3) to characterize its CA inhibitory activity, and confirmed that it is a very weak inhibitor, unlike 1 or 2. In a CO₂ hydration assay, 3 behaved as a very weak, partial inhibitor of CA-II and CA-I. We conclude that topiramate (1), zonisamide (2), and sulfamide 3 do not require extended exposure to human CA-I or CA-II to manifest full inhibitory activity (4-NPA assay).

Keywords: Zonisamide, topiramate, carbonic anhydrase, inhibition, sulfamide, preincubation

Introduction

Carbonic anhydrases (CAs) are important enzymes that have attracted considerable interest [1,2]. Given the roles of CAs in many physiological and pathophysiological processes, CA inhibitors can serve as useful therapeutic agents for treating numerous diseases, including glaucoma, cancer, and obesity [2–4]. Topiramate (TPM; 1) and zonisamide (ZNS; 2) are widely marketed antiepileptic drugs that evince some CA inhibitory actions. Although these drugs have relatively little structural similarity, they do possess analogous sulfamate and sulfonamide moieties, respectively. Additionally, these drugs share several pharmacological properties in that they inhibit some types of neuronal voltage-gated Na⁺ and Ca²⁺ channels [5], and certain isoforms of CA[2]. When their CA inhibitory activity was first identified in the 1980's, these compounds were viewed as comparatively weak inhibitors (e.g., relative to acetazolamide),

with K_i values for ZNS in the range of 4–12 μM[6–8] and K_i values for TPM in the range of 1–120 μM[9–11] (for CA activity in erythrocytes, brain, or kidney; in various species). As knowledge about the various CA isozymes developed, and as CA isozymes became available in purified forms, it has become possible to establish more accurate K_i values for each CA isozyme[1–4].

Recently, we have conducted studies involving four distinct assay procedures to determine K_i or K_d values for the interaction of TPM with human CA-II and CA-I[12–15]. The K_i or K_d values from the four procedures consistently ranged from 0.3–0.6 μM for CA-II and 90–140 μM for CA-I. In contrast, another research group has reported much more potent inhibition of human CA-II and CA-I by TPM, with K_i values of 0.005–0.010 μM and 0.25 μM, respectively[16,17]. This apparent discrepancy in K_i values of approximately 60-fold (CA-II) and 400-fold (CA-I) provided an impetus for us to seek independent

Correspondence: B. E. Maryanoff, Ph.D., Johnson & Johnson Pharmaceutical Research & Development, Spring House, Pennsylvania 19477-0776, USA. Tel.: + 1-215-628-5530. Fax: + 1-215-628-4985. E-mail: bmaryano@prdus.jnj.com

[†]Tel.: + 1-215-646-5184. E-mail: richardshank@comcast.net

[‡]Tel.: + 1-215-628-5590. Fax: + 1-215-628-4985. E-mail: vsmithsw@prdus.jnj.com

verification through a direct thermodynamic method (ThermoFluor®), which afforded a K_d value of $0.3 \mu\text{M}$ for TPM[14]. Since the marked differences in K_i (or K_d) values between the two research groups have yet to be adequately resolved, we became intrigued by the possibility that slow-tight binding kinetics might play a role. Indeed, a recent publication reports slow-tight binding behavior for ZNS with CA-II[18]. According to De Simone et al.[18], establishing a true K_i for ZNS as an inhibitor of CA-II requires one-hour preincubation of ZNS with CA-II prior to initiation of the catalytic reaction. More specifically, preincubation of ZNS for 15 minutes with CA-II yielded a K_i of $10.3 \mu\text{M}$; however, preincubation for 60 minutes yielded a much more potent K_i of $\sim 35 \text{ nM}$ [18]. We have performed experiments to replicate these findings with ZNS, as well as to determine if the K_i values for TPM and its sulfamide cognate (**3**; JNJ-17065984) would require such an extended preincubation period. In contradistinction, we found that preincubation of up to one hour had essentially no effect on the K_i value for ZNS with either CA-II or CA-I. The same behavior was observed for TPM and sulfamide **3**. Additionally, the present study confirms our prior observations that sulfamide **3** is an exceedingly weak inhibitor of CA-II and CA-I[13–15].

Materials and methods

Test compounds

Topiramate (**1**) and its sulfamide cognate **3** were synthesized as described earlier[13]. Zonisamide (**2**) and 4-nitrophenylacetate (4-NPA) were purchased from Sigma Chemical Co. (St. Louis, MO). Test compounds were dissolved in DMSO at 100 mM and diluted in purified water just prior to the kinetics measurements.

Enzyme reactions

In most of the experiments we used 4-nitrophenylacetate (4-NPA) as the substrate for enzyme catalysis (Figures 1–4). 4-NPA (colorless) is hydrolyzed to 4-nitrophenol (yellow) and acetate, and the reaction is monitored by the emergence of color[19,20]. A detailed description of the assays used in this study was reported previously[12,15]. The incubation medium was buffered with HEPES (10 mM) and the pH was adjusted to 7.3 with Tris. For each sample, the reaction volume included 70 μL of the HEPES–Tris solution, 10 μL of an aqueous solution of test compound (10 μL of H_2O for catalyzed control sample), 10 μL of a freshly prepared aqueous solution of purified human erythrocyte CA-II (~ 0.02 to $0.1 \mu\text{g}$) or CA-I (~ 0.1 to $0.5 \mu\text{g}$) (10 μL of H_2O for uncatalyzed reaction samples), and 10 μL of a solution in which

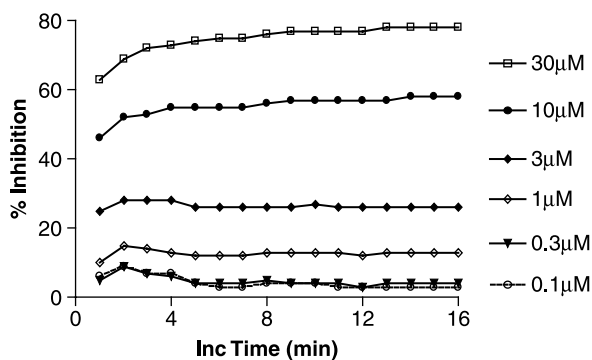


Figure 1. Graphic representation of the percent inhibition of CA-II by ZNS as a function of the concentration of ZNS and the incubation (Inc) time after initiation of the enzymatic reaction (4-NPA hydrolysis assay). The increase in the optical density was monitored at 1-minute intervals. The data derive from one experiment performed in quadruplicate.

4-NPA was dissolved in DMSO, then diluted 10-fold with absolute ethanol. The concentration of 4-NPA in the reaction medium was 3 mM. The reaction was performed at 21–23°C in a 96-well plate. The reaction was initiated by adding 4-NPA stock solution either within a few minutes, or at 60 min, after addition of the enzyme. In each experiment, test compounds were evaluated in quadruplicate. Immediately after initiating the reaction, the plate was placed in a SPECTRAMax 384 spectrophotometer (Molecular Devices, Sunnyvale, CA) and the formation of reaction product was monitored at 1-min intervals for 15–20 min at 400 nm. The concentration of 4-NPA was at least 25-fold lower than the K_m of 4-NPA as a CA-II substrate[19,20]. According to the Cheng–Prusoff equation[21], under these conditions the IC_{50} and K_i values differ by less than 3%, and they are regarded as virtually synonymous in this study.

Data analysis and calculation of K_i values

The relative difference in the spectrophotometric optical density between the uninhibited CA-catalyzed reaction and the uncatalyzed reaction (nonspecific activity) was calculated and assigned a value of 100. The data for each test compound were referenced to the two sets of control data, from which the percent inhibition of CA activity was calculated. Replicates for each experiment were averaged. Each compound was tested in two or three separate experiments at six concentrations. From the concentration–inhibition data, IC_{50} (K_i) values were calculated by using PRISM 4 (Graphpad, San Diego, CA). Two curve-fit equations were used to analyze the data. One equation assumed single site normal competition saturation kinetics $[\%I = (100 - 0)/(1 + 10^{(\text{Log}X - \text{Log}EC_{50})})]$; where 100 and 0 represent the maximum and minimum possible % inhibition (I), respectively, and X is the inhibitor concentration) and the other allowed

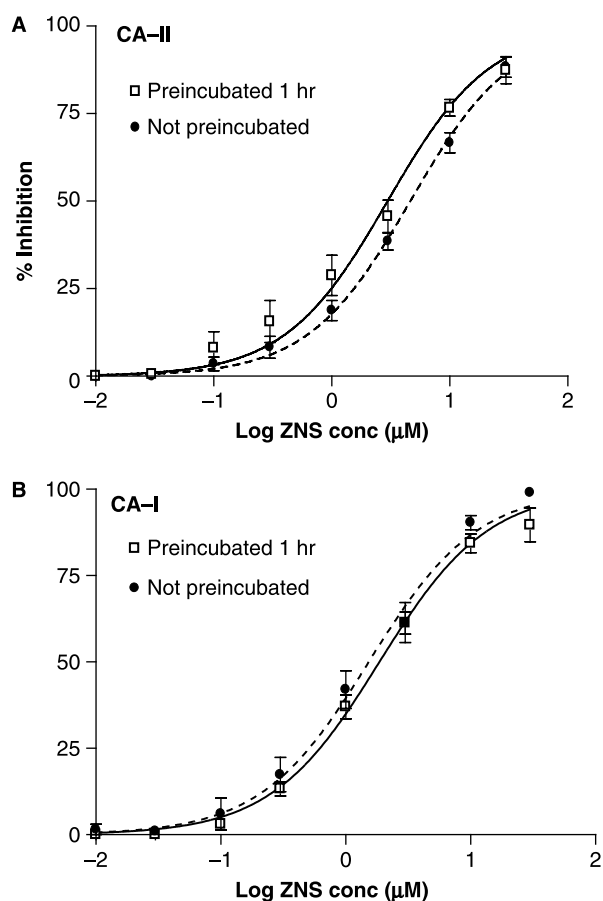


Figure 2. Concentration–inhibition graphs illustrating the effect of preincubation on the inhibition of CA-II and CA-I by ZNS (4-NPA assay). The data are the mean (\pm SEM) of 4 or 5 experiments, each of which was performed in quadruplicate. The curve-fit analysis of the data assumed normal saturation and one-site competition. Filled circles are paired with the dashed line.

the saturation kinetics to vary and generate a variable Hill slope with maximum inhibition of $\leq 100\%$ [$\%I = (100 - 0)/(1 + 10^{-(\text{LogEC}_{50} - \text{LogX}) \cdot B})$]; where 100, 0, I, and X are the same as above, and B is the Hill slope].

Results

At the outset, we conducted an experiment with ZNS involving CA-II-catalyzed 4-NPA hydrolysis to ascertain the rate at which the CA-II inhibitory effect approached equilibrium when there was no prior exposure of the enzyme to ZNS (Figure 1). Our results indicate that the binding equilibrium is achieved within 15 minutes *even in the absence of preincubation*.

In subsequent experiments, we determined the effect of 60-minute preincubation of ZNS with CA-II or CA-I on the calculated K_i (Figure 2). For one set of samples, the hydrolysis of 4-NPA was initiated within a few minutes after ZNS and the enzyme (CA-II or CA-I) were combined in the incubation medium, after which the samples were “read” every minute over

a 20-minute period. The raw data from this 20-minute experiment were used to calculate the K_i values. Another set of samples was treated identically except that there was a 60-minute delay prior to initiating the hydrolysis of 4-NPA. The concentration–inhibition data for each of the four sets of samples modeled well to the equation describing normal saturation for one-site competition kinetics (Figure 2). The K_i (95% confidence interval, 95% CI) values from this analysis for inhibition of CA-II were 2.98 (2.30–3.85) μM for the preincubated samples and 4.64 (4.06–5.30) μM for the samples that were not preincubated (Figure 2A). The corresponding K_i (95% CI) values for the inhibition of CA-I were 1.86 (1.62–2.15) μM and 1.53 (1.22–1.92) μM for the preincubated and non-preincubated samples, respectively (Figure 2B).

A similar set of experiments with CA-II or CA-I was conducted for TPM to determine the effect of preincubation on the K_i value (Figure 3). The concentration–inhibition data were subjected to a curve-fit analysis assuming one-site competition with normal saturation kinetics, as well as a curve-fit

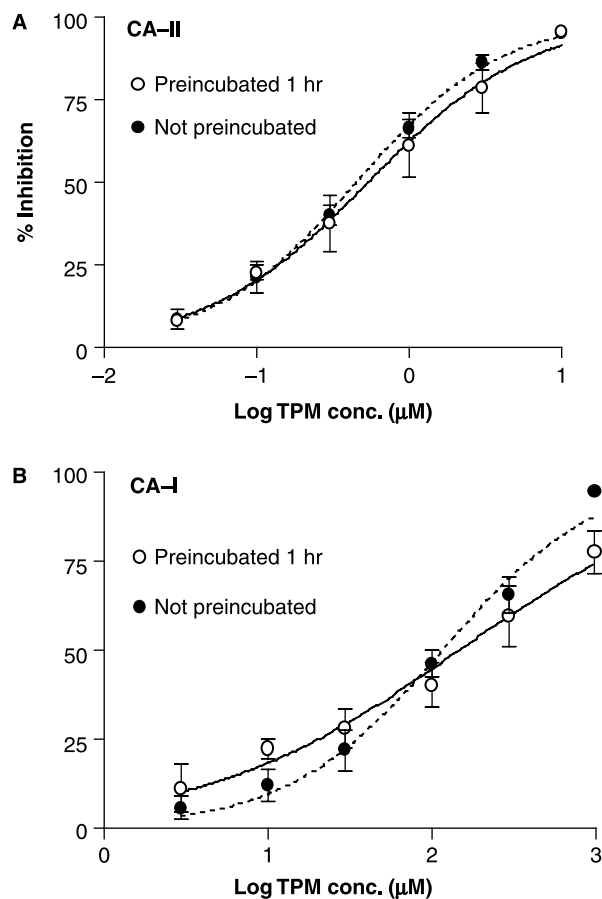


Figure 3. Concentration–inhibition graphs showing the effect of preincubation on the inhibition of CA-II and CA-I by TPM (4-NPA assay). The data are the mean (\pm SEM) of 4 or 5 experiments (each done in quadruplicate). The curve-fit analysis of the data was performed with a sigmoidal saturation equation that allows for a variable slope. Filled circles are paired with the dashed line.

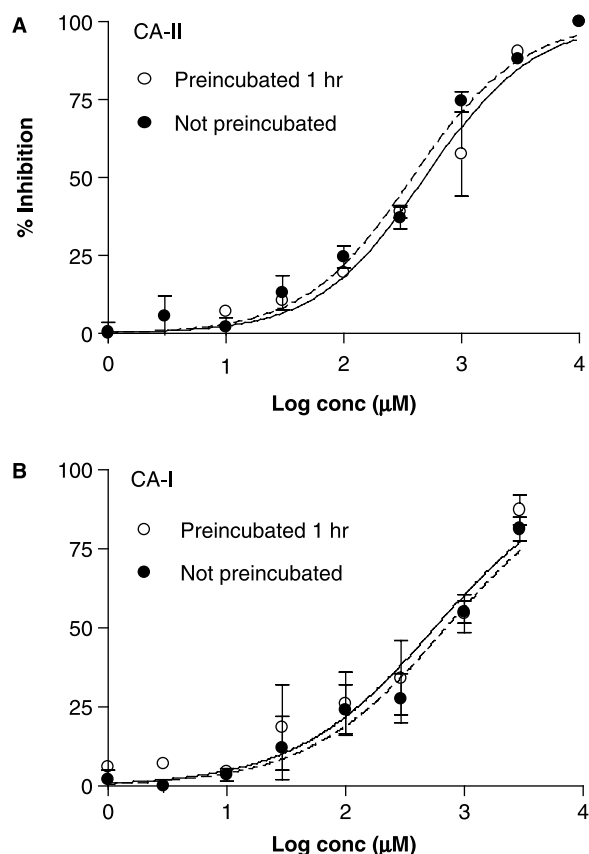


Figure 4. Concentration–inhibition graphs illustrating the effect of preincubation on the inhibition of CA-II and CA-I by **3** (4-NPA assay). The data are the mean (\pm SEM) of 3 or 6 experiments (each done in quadruplicate). The curve-fit analysis of the data was performed using a sigmoid saturation equation that allows for a variable slope. Filled circles are paired with the dashed line.

analysis that used a sigmoidal dose-response with a variable slope. From this analysis, the K_i (95% CI) values for the inhibition of CA-II were 0.54 (0.38–0.76) μM for the preincubated samples and 0.45 (0.38–0.53) μM for the non-preincubated samples (Figure 3A). (The K_i values derived from the two curve-fitting procedures differed by less than 1%.) Since the inhibition of CA-I by the preincubated set of samples exhibited a low Hill slope (0.55), we elected to use a curve-fit analysis with a variable slope (Figure 3B). The corresponding K_i (95% CI) values for the inhibition of CA-I were 147 (93–234) and 117 (92–150) μM for the preincubated and non-preincubated experiments, respectively (Figure 3B).

We also examined the sulfamide cognate of TPM (**3**; JNJ-17065984) in this manner, since the K_i values for **3** reported by us and Supuran's group have differed widely (as mentioned above). For **3**, we have obtained consistent K_i values of 340, 456, and 650 μM in three independent experiments, as well as a K_d value of 25 μM [13–15]. On the other hand, Supuran and coworkers reported a K_i value of 2 μM [17]. Could this

discrepancy in the K_i values be attributable to differences in preincubation time? To address this issue, we not only performed a set of experiments similar to those performed for ZNS and TPM, but also an experiment involving CA-catalyzed hydration of CO_2 , which is described in detail elsewhere[13,15]. The concentration–inhibition data for the 4-NPA hydrolysis assay with **3** were subjected to the two curve-fit analyses, as described above for TPM. The inhibition of both CA-II and CA-I exhibited relatively low Hill slopes (0.74). For this reason, the curve-fit analysis using the variable slope was chosen again. The K_i (95% CI) values for the inhibition of CA-II by **3** were 382 (315–464) μM for the preincubated samples and 489 (367–652) μM for the non-preincubated samples (Figure 4A). The corresponding K_i (95% CI) values for the inhibition of CA-I by **3** were 569 (347–933) and 707 (506–989) μM for the preincubated and non-preincubated samples, respectively (Figure 4B).

In a previous study of **3** with the CO_2 hydration procedure, we found it to be a weak, partial inhibitor of CA-II[15]. To substantiate that observation, we now performed experiments with expanded concentration ranges: 0.1 μM to 10 mM for CA-II (Figure 5A) and 10 μM to 10 mM for CA-I (Figure 5B). With both enzymes, the inhibitory activity at the 10 mM concentration of **3** was far below the maximum possible inhibition of 100%. A PRISM curve-fit analysis indicated that the maximum inhibition of CA-II and CA-I by **3** was just $\sim 50\%$ and $\sim 33\%$, respectively. The IC_{50} values obtained by curve-fit analysis were converted to K_i values by using the Cheng–Prusoff equation[21]. The K_i (95% CI) value for inhibition of CA-II was 243 (3.3–17800) μM , with a low Hill slope of 0.48 ± 0.24 (SEM) and maximum % inhibition of 50 ± 25 . The exceptionally wide 95% CI for the K_i is attributable to the very shallow concentration–inhibition curve (Figure 5A), which turned out to cause rather large standard errors. The K_i value for inhibition of CA-I was 48.7 (21.2–112) μM , with a Hill slope of 2.0 ± 1.4 (SEM) and a maximum % inhibition of 33 ± 5 μM . Evidently, the CO_2 hydration assay has limitations for assessing very weak inhibitors such as **3**.

Discussion

The results of our investigation indicate that the interaction of ZNS with CA-II or CA-I reached equilibrium within a 15-minute period. Also, extending the exposure time prior to initiating the catalytic reaction with CA-II or CA-I had little or no effect on the K_i values generated with the 4-NPA hydrolysis assay. In the same vein, extending the exposure of TPM or sulfamide **3** with either isozyme to one hour prior to adding 4-NPA had little or no effect on the K_i values (relative to those obtained at 15 minutes).

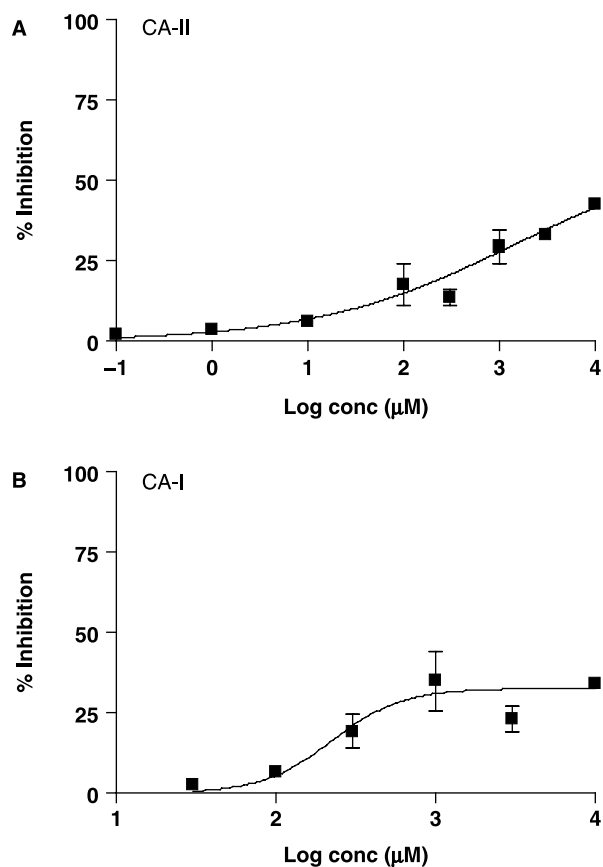


Figure 5. Concentration–inhibition graphs depicting the inhibitory effect of **3** on the activity of human CA-II and CA-I as determined with a CO_2 hydration assay (described in detail previously[15]), instead of the 4-NPA assay. The data are the mean (\pm SEM) of 2–6 experiments (each performed in triplicate). The curve-fit analysis of the data was performed with a sigmoidal saturation equation that allowed for a variable slope and variable maximum % inhibition. The K_i was 2.4-fold and 4.6-fold lower than the IC_{50} for CA-II and CA-I, respectively.

On the basis of these results, extended preincubation of the inhibitor with the enzyme cannot be responsible for the disparate K_i values reported by us[12–15] and Supuran’s group[16,17]. Moreover, the K_i values for the inhibition of CA-II by **3** of 380 μM (preincubated) and 490 μM (non-preincubated) are consistent with our prior observations of exceedingly weak inhibition of CA-II[13–15]. In fact, we were the first to note that sulfamides appear to be much weaker inhibitors of CA-II than their sulfamate congeners[13]. Later, we substantiated this point with an independent assay based on direct binding to CA-II (ThermoFluor® method)[22], which yielded thermodynamically-based K_d values[14]. For **3**, the ThermoFluor technique afforded a K_d value of 25 μM , which is 50-fold weaker than the K_d value for TPM[14]. The weaker potency of a sulfamide relative to its sulfamate cognate may be ascribed to the former being much less acidic, with a larger $\text{p}K_a$ value by nearly 2 units[14]. This ~ 100 -fold lower acidity for the sulfamide would mean that it has a much lower population of the

anionic form required for binding to Zn(II) in the CA active site. An alternative steric explanation for **3** being a weaker inhibitor of CA-II than TPM (**1**) was proposed by Winum et al. from an examination of the X-ray co-crystals of **1**·CA-II and **3**·CA-II[17].

The K_d (ThermoFluor) and K_i (CO_2 hydration assay) values for an inhibitor compound, have generally shown reasonable consistency (≤ 3 -fold difference)[14]. However, there was divergence from this picture in the case of sulfamide **3**, for which the K_i was ~ 25 -fold higher than the K_d ($K_i = 650 \mu\text{M}$ vs. $K_d = 25 \mu\text{M}$). In a subsequent study, we compared sulfamates to sulfamides as inhibitors of CA-II and CA-I, and obtained K_i values by utilizing both the CO_2 hydration and 4-NPA hydrolysis assays[15]. Thus, we encountered very shallow concentration–inhibition curves in the CO_2 hydration assay with several sulfamides, reflecting characteristics of partial and/or mixed inhibition. By contrast, the same sulfamides demonstrated full inhibition in the 4-NPA hydrolysis assay. To investigate this issue further for sulfamide **3**, we expanded the concentration conditions to encompass a 100,000-fold range in the current study (Figure 5A). It was hoped that such a broad range would establish the maximum inhibition of CA-II more accurately and thereby furnish a more accurate K_i value. However, despite this expanded concentration range, the K_i value of 243 μM from the PRISM curve-fit analysis had an extremely wide 95% confidence interval of 3.3–17800. Consequently, in the CO_2 hydration assay sulfamide **3** is a very weak, partial inhibitor of CA-II, such that it is difficult to establish a reasonably accurate K_i value for **3**.

In the various assays that have been employed to test compounds for inhibition of CA isozymes, the biochemical principles and experimental conditions can vary greatly. For example, assays based on the hydration of CO_2 are typically performed at lower temperatures (0–5°C) to minimize uncatalyzed hydration of CO_2 , whereas the ThermoFluor assay is performed at higher temperatures (50–60°C) to monitor the molecular folding state of the enzyme. The temperature differences in these two assays are likely to have some influence on the determined inhibitor–enzyme affinity values. The operative question is: What would be considered reasonable differences in K_i or K_d values generated by different assays performed in various laboratories? In this respect, the following issues come into play: (1) the intrinsic properties of the assay procedure, including the design of the experiment (e.g., number of concentrations and replicates for each inhibitor; the concentration(s) of the substrate); (2) the method used to analyze the data; and (3) normal experimental error that is inherent for each assay procedure. Because of the numerous variables involved in generating the raw data and the various ways of calculating K_i or K_d values, differences of as much

as 3-fold would be standard and acceptable. However, when the differences exceed 5-fold, and especially 10-fold, it becomes important to identify and understand the cause(s). Our current study was performed with this aspect in mind. First, we sought to identify the source of the 26-fold difference between the K_d and K_i values for sulfamide **3** obtained in our own laboratories[14], as described above. Second, we sought to resolve the issue related to the 60-fold to ~400-fold difference between our reported K_i and K_d values for TPM (0.3–0.6 μ M for human CA-II; 90–150 μ M for human CA-I)[12–15] and the K_i values reported by Supuran and coworkers (0.005–0.010 μ M for human CA-II; 0.25 μ M for human CA-I)[16,17]. In the present study, our CA-II K_i value for **3** of 380–490 μ M still differs by ca. 200-fold from the CA-II K_i value of 2 μ M reported by Winum et al[17].

Winum et al.[17] insinuated that the CA inhibition data produced in our published studies may be flawed due to the presence of Zn^{++} chelators in the incubation medium, without proffering any evidence, experimental or otherwise, that there could be such an effect. Perhaps, their suggestion may be based on the notion that dithiothreitol (DTT) is included in some CO_2 hydration-based assays[23], and that DTT at very high concentrations can bind to CA-bound Zn, thereby attenuating the CA activity[24]. We have performed experiments without DTT in the reaction medium, or with DTT at the concentration recommended by Cammer et al.[23], and found no differential effects on the activity of CA-II or CA-I (control experiments; not shown). Additionally, with or without DTT, there were no differences in the potencies of CA-II or CA-I inhibition for TPM or the reference inhibitor acetazolamide (control experiments; not shown). Along this line, it is important to note that EDTA, a well-known Zn^{++} chelator, actually *enhances* the affinity of various sulfonamide inhibitor ligands for CA-II and CA-I; it does not diminish their affinity[22]. Consequently, a Zn^{++} chelator is highly unlikely to be responsible for the higher K_i and K_d values that we have reported for TPM[12–15].

Time and again, through several independent lines of investigation, we have demonstrated the key points of our claims. From the work described herein, we firmly corroborate the K_i values for zonisamide (**2**), topiramate (**1**), and the sulfamide cognate of topiramate (**3**; JNJ-17065984). Moreover, we conclude that zonisamide, topiramate, and sulfamide **3** do not require extended preincubation to express full

activity as inhibitors of CA-II and CA-I. Finally, we wish to underscore that sulfamide **3** is an exceedingly weak inhibitor of both CA isozymes.

References

- [1] Wistrand PJ. In: Chegwidden WR, Carter ND, Edwards YH, editors. *The carbonic anhydrases: New horizons*. Basel: Birkhäuser Verlag; 2000. p 597–609.
- [2] Scozzafava A, Mastrolorenzo A, Supuran CT. *Expert Opin Ther Pat* 2004;14:667–702.
- [3] Maren TH. In: Chegwidden WR, Carter ND, Edwards YH, editors. *The carbonic anhydrases: New horizons*. Basel: Birkhäuser Verlag; 2000. p 425–436.
- [4] Parkkila S, Parkkila A-K, Kivela J. In: Supuran CT, Scozzafava A, Conway J, editors. *Carbonic anhydrase: Its inhibitors and activators*. Boca Raton, FL: CRC Press; 2004. p 283–301.
- [5] Rogawski MA, Löscher W. *Nat Rev Neurosci* 2004;5:553–564.
- [6] Matsumoto K, Miyazaki H, Fujii T, Hashimoto M. *Chem Pharm Bull* 1989;37:1913–1915.
- [7] Masuda Y, Karasawa T. *Arzneim-Forsch* 1993;43:416–418.
- [8] Masuda Y, Noguchi H, Karasawa T. *Arzneim.-Forsch* 1994;44:267–269.
- [9] Maryanoff BE, Nortey SO, Gardocki JF, Shank RP, Dodgson SP. *J Med Chem* 1987;30:880–887.
- [10] Shank RP, Gardocki JF, Vaught JL, Davis CB, Schupsky JJ, Raffa RB, Dodgson SJ, Nortey SO, Maryanoff BE. *Epilepsia* 1994;35:450–460.
- [11] Dodgson SJ, Shank RP, Maryanoff BE. *Epilepsia* 2000;41 (Suppl. 1), S35–S39.
- [12] Shank RP, Doose DR, Streeter AJ, Bialer M. *Epilepsy Res* 2005;63:103–112.
- [13] Maryanoff BE, McComsey DF, Costanzo MJ, Hochman C, Smith-Swintosky V, Shank RP. *J Med Chem* 2005;48: 1941–1947.
- [14] Klinger AL, McComsey DF, Smith-Swintosky V, Shank RP, Maryanoff BE. *J Med Chem* 2006;49:3496–3500.
- [15] Shank RP, McComsey DF, Smith-Swintosky VL, Maryanoff BE. *Chem Biol Drug Design* 2006;68:113–119.
- [16] Casini A, Antel J, Abbate F, Scozzafava A, David S, Waldeck H, Schäfer S, Supuran CT. *Bioorg Med Chem Lett* 2003;13: 841–845.
- [17] Winum J-Y, Temperini C, Cheikh KE, Innocenti A, Vullo D, Ciattini S, Montero J-L, Scozzafava A, Supuran CT. *J Med Chem* 2006;49:7024–7031.
- [18] De Simone G, Di Fiore A, Menchise V, Pedone C, Antel J, Casini A, Scozzafava A, Wurlb M, Supuran CT. *Bioorg Med Chem Lett* 2005;15:2315–2320.
- [19] Thorslund A, Lindskog S. *Eur J Biochem* 1967;3:117–123.
- [20] Pocker Y, Stone JT. *Biochemistry* 1967;6:668–678.
- [21] Cheng Y, Prusoff WH. *Biochem Pharmacol* 1973;22: 3099–3108.
- [22] Matulis D, Kranz JK, Salemm FR, Todd MJ. *Biochemistry* 2005;44:5258–5266.
- [23] Cammer W, Fredman T, Rose AL, Norton WT. *J Neurochem* 1976;27:165–171.
- [24] Husic HD, Hsieh S, Berrier AL. *Biochim Biophys Acta* 1991; 1078:35–42.

Copyright of *Journal of Enzyme Inhibition & Medicinal Chemistry* is the property of Taylor & Francis Ltd and its content may not be copied or emailed to multiple sites or posted to a listserv without the copyright holder's express written permission. However, users may print, download, or email articles for individual use.